

TOTAL SYNTHESIS OF NEOMYCIN B†

Sir:

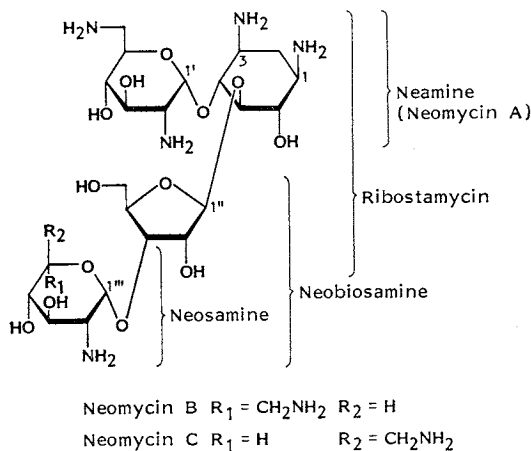
Neomycin B is a main component of the neomycin complex which was independently discovered by UMEZAWA *et al.*¹⁾ and WAKSMAN and LECHEVALIER²⁾. Neomycin group is a representative of pseudo-tetrasaccharide antibiotics, which provide some fascinating problems in carbohydrate chemistry^{3,4)}. Complete structures and absolute stereochemistry of neomycins were elucidated by HICHENS and RINEHART, Jr.⁵⁾. Total synthesis of neomycin C, another component of the neomycin complex, has been achieved by UMEZAWA and NISHIMURA⁶⁾ and UMEZAWA *et al.*⁷⁾. Neamine (neomycin A), a pseudo-disaccharide portion common to neomycins, was synthesized also by UMEZAWA *et al.* first⁸⁾ through conversion of paromamine and later⁹⁾ by glycosylation of the aminocyclitol component. On the other hand, because of the difficulty associated with glycosylation by 2,6-diamino-2,6-dideoxy-L-idose (neosamine B) which distinguishes neomycin B from C, total synthesis of neomycin B has not been successful¹⁰⁾. In this communication we wish to report the first total synthesis of neomycin B.

In order to solve the above-mentioned problem, we have reported the synthesis of 3-O-acetyl-2,6-diazido-4-O-benzyl-2,6-dideoxy-L-idopyranosyl chloride (1)¹¹⁾ as a key intermediate for the preparation of 1,2-*cis*(β -L-) linked neo-

samine B. The present approach to neomycin B is characterized by stepwise glycosylation reactions: 1) Glycosylation of the 3-OH of a protected ribose (2)¹²⁾ with glycosyl chloride (1) to yield a masked disaccharide (3) corresponding to neobiosamine B, 2) glycosylation of the 5-OH of a selectively protected neamine (10) with the disaccharide portion to yield a masked pseudo-tetrasaccharide (11), and subsequent transformation of 11 into neomycin B.

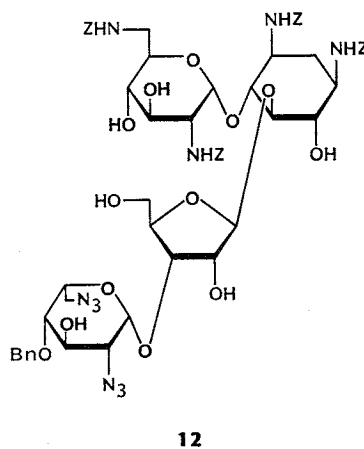
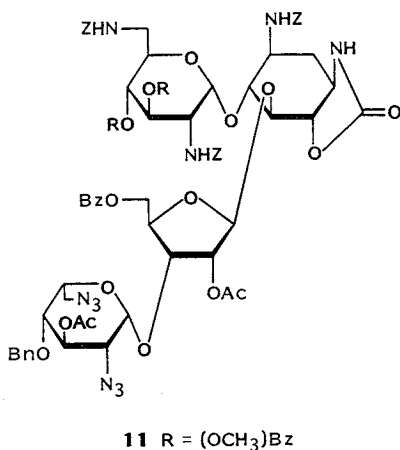
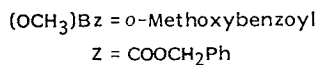
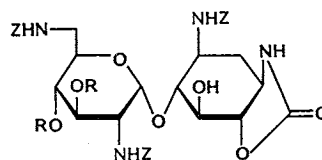
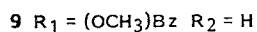
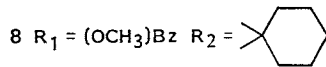
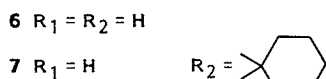
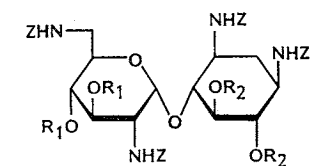
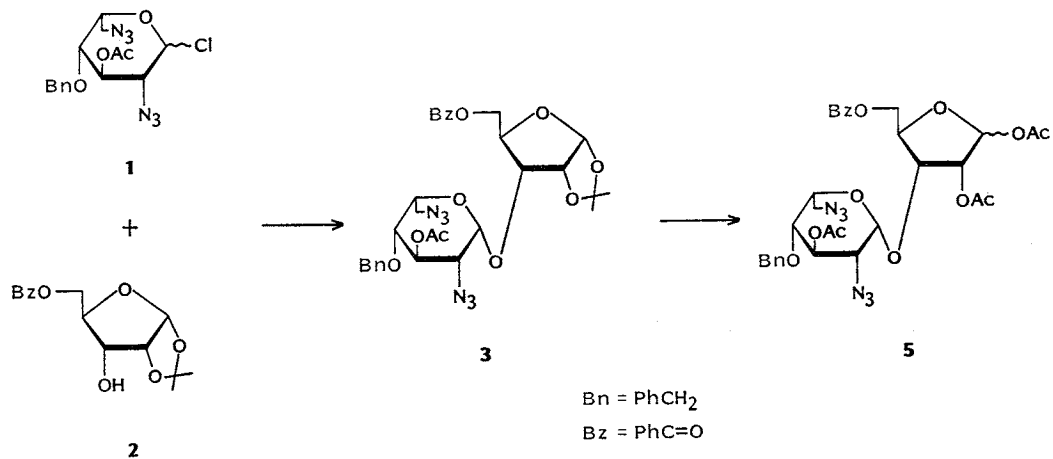
Reaction of 1 with 2 was performed under modified KOENIGS-KNORR condition (mercuric cyanide, mercuric bromide, Molecular sieves 4A, CH₂Cl₂, room temp overnight), yielding the desired β -L-connected disaccharide 3 in 70% yield, based on 2, and the α -L anomer (4, 7%). The anomeric configurations were assigned on the basis of the more dextro-rotating specific rotation^{††} of 3 (+139°; 4 +28°) and their NMR spectra¹³⁾. ¹H NMR (CDCl₃, 250 MHz); 3: δ 5.11 (d, $J_{1',2'}$ =2.0 Hz, 1'-H); 4: δ 4.87 (d, $J_{1',2'}$ =3.4 Hz, 1'-H). ¹³C NMR (CDCl₃, 62.9 MHz); 3: δ 97.5 (C-1'); 4: δ 100.1 (C-1'). Deisopropylideneation (AcOH-2 N HCl, 2:1) of 3 followed by conventional acetylation (Ac₂O-pyridine) gave an anomeric mixture (5, α : β ~ 1:3) of glycosyl acetates, 92%; [α]_D +82°, a neobiosamine B equivalent duly masked and functionalized for the coupling with protected neamine.

A new neamine derivative, in which all the functional groups except the 5-hydroxyl are protected, was prepared. Selective cyclohexylideneation of tetrakis(*N*-benzyloxycarbonyl)-neamine (6)¹⁴⁾ by a modified literature procedure gave 7¹⁵⁾, the two hydroxyl groups of which were then acylated with *o*-methoxybenzoyl group to give a fully protected neamine 8, 72% from 6; [α]_D +22.5°. Decyclohexylideneation (AcOH-H₂O-THF, 3:1:1) gave the diol 9, 98%; [α]_D +59°. Treatment of 9 with sodium hydride in DMF in the presence of benzyltriethylammonium chloride gave cyclic 1,6-carbamate 10, 78%; [α]_D +80°; ¹H NMR (CDCl₃+D₂O, 250 MHz) δ 1.55 (1H, q, J =~12 Hz, 2-H_{ax}), 2.15 (1H, m, 2-H_{eq}), 3.65 and 3.68 (each 3H, s, OCH₃). Addition of the quaternary ammonium salt was found to improve the yield.



† Presented in part at the 54th National Meeting of the Chemical Society of Japan, Tokyo, April 1987, Abstr. 4111L16.

†† Optical rotations were measured, unless otherwise noted, at 20~22°C for solutions (c 0.7~1.1) in CHCl₃. All new compounds gave satisfactory elemental analyses.



Coupling of the two portions was best effected by use of trimethylsilyl triflate (TMSOTf)¹⁶. Reaction of 10 with 5 (1.5 equivalent to 10) in the presence of TMSOTf (benzene, room temp

4 hours) gave pseudo-tetrasaccharide 11 as a sole isolable product, 60%; [α]_D +55°. Treatment of 11 with sodium benzyl alcoholate gave partially deprotected 12, 85%; [α]_D +40°.

Table 1. Antibacterial spectra of synthetic and natural neomycin B.

Test organisms*	MIC ($\mu\text{g/ml}$)	
	Synthetic	Natural
<i>Staphylococcus aureus</i> FDA 209P	0.78	0.78
<i>Micrococcus luteus</i> PCI 1001	1.56	0.78
<i>Bacillus subtilis</i> NRRL B-558	0.20	0.39
<i>Escherichia coli</i> K-12	0.78	0.39
<i>E. coli</i> K-12 ML1629	100	100
<i>E. coli</i> K-12 LA 290 R 55	1.56	1.56
<i>E. coli</i> K-12 LA 290 R 56	1.56	0.78
<i>E. coli</i> K-12 W 677	0.78	0.39
<i>E. coli</i> K-12 JR 66/W 677	100	100
<i>Klebsiella pneumoniae</i> PCI 602	0.78	0.78
<i>K. pneumoniae</i> type 22 #3038	100	100
<i>Mycobacterium smegmatis</i> ATCC 607**	0.78	0.78

* Agar dilution streak method (nutrient agar, 37°C, 17 hours).

** 48 hours.

Finally catalytic hydrogenation (4 kg/cm² H₂, 10% Pd-C) of **12** gave neomycin B, 65% (as free base). ¹H¹⁷) and ¹³C NMR¹⁸⁾ spectra of the free base (in D₂O) and the specific rotation¹⁹⁾ of the hydrochloride [$+54^\circ$ (c 1.1, H₂O)] agreed well with those reported. The identity of the synthetic and natural specimens was further established by the antibacterial spectra as shown in Table 1.

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